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(54) Title: <b>HYDROXYAPATITE-ANTIGEN CONJUGATES AND METHODS FOR GENERATING A POLY-IG IMMUNE RESPONSE</b>			
(57) Abstract			
<p>A method for generating antigen-sensitized Ig-A-producing lymphoblasts in a mammal, using an immunogen comprising an antigen or antigen mixture in association with hydroxylated calcium phosphate (hydroxyapatite) is administered to a mucosal surface of the mammal.</p>			

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HYDROXYAPATITE-ANTIGEN CONJUGATES  
AND METHODS FOR GENERATING A  
POLY-Ig IMMUNE RESPONSE

5

Background of the Invention

This invention was made in part with funding from the United States Government, and the U.S. Government has certain rights in the invention.

This invention relates to the general fields of 10 passive mucosal immune protection, and of poly-Ig immune reagents and techniques. We use the term poly-Ig to refer to the polymeric classes of antibodies--i.e. IgA and IgM. IgM antibodies are generally produced at an early stage of the immune response and are not an important factor in protective mucosal immunity. Thus, 15 the invention generally refers to polymeric Ig antibodies, and the usual and preferred antibodies for all aspects of the invention are IgA class antibodies, which normally are secreted in dimeric form and, to a 20 lesser degree, as higher IgA polymers.

Many pathogenic bacteria and viruses initially gain entry into the body by crossing the cellular linings (epithelia) of the gastrointestinal, respiratory, or genital tracts. A specialized class of antibodies, IgA 25 antibodies, protects these surfaces. IgA antibodies are dimeric or polymeric molecules produced by cells located in the tissues under the epithelial surfaces. They are transported by epithelial cells into mucosal secretions, where they cross-link or coat pathogens that have not yet 30 entered the body, preventing the pathogens from contacting and adhering to epithelial cells. Thus, IgA antibodies operate on pathogens that are outside the body, and they protect by preventing entry into the body across epithelial surfaces.

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The naturally occurring IgA response is triggered by antigen delivery to mucosal surfaces. The antigen enters the body through specific sampling sites (termed microfold or M cells) that effect transepithelial antigen  
5 transport to areas of the mucosal lining containing specialized, organized collections of the cells of the mucosal immune system. More specifically, as shown in Fig. 1, antigens A (shown as filled dots) in lumen 1 bind the luminal surface of M cells at site 2. The antigens  
10 are internalized and transcytosed at 3 to be released in the intra-epithelial pocket 4 containing lymphoid cells L (B and T cells) and antigen-processing/presenting cells such as macrophage cells (M).

IgA antibodies in a naturally immunized host are  
15 transported into secretions by binding to a specific receptor (called the poly-Ig receptor) on the basal (interior) surfaces of epithelial and glandular cells throughout the respiratory and digestive systems, the genital tract, and the mammary glands. See Solari and  
20 Kraehenbuhl, "Receptor-Mediated Transepithelial Transport of Polymeric Immunoglobulins", pp.269-298 in The Mammary Gland, Nelville and Daniel Eds., Plenum Publishing, Cambridge (1987); Mestecky (1987) J. Clin. Immunol. 7:265-276. Receptor-IgA complexes are transported across  
25 these cells and exocytosed onto luminal (exterior) cell surfaces where the receptor is enzymatically cleaved, releasing IgA into secretions along with a receptor fragment called secretory component (SC). See Mostov et al. (1980) Proc. Nat'l Acad. Sci. U.S.A. 77:7257-7261;  
30 Solari, R. and Kraehenbuhl, J.P. Cell 36:61-71 (1984); Kuhn and Kraehenbuhl, J. Biol. Chem. 256:12490-12495(1981). It is reported that secretory component reduces proteolytic degradation of IgA. Lindh,  
J., J. Immunol. 114:284-286 (1975); Brown, Neucomb,  
35 Ishizaka, J. Clin. Invest. 49:1374 (1974).

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In general, existing immunization strategies which involve injection of antigens evoke production of the IgG class of antibodies that circulate systemically and neutralize pathogens after they have entered the body.

5 Injection of antigens does not generally evoke a substantial IgA response.

Efforts to take advantage of IgA protection at mucosal barriers involve oral immunization, either for active protection of the immunized mammal or for passive 10 protection of another mammal using mucosal secretion of the immunized mammal. Glass et al., New Eng. J. Med., 308:1389-1392 (1983); Fubara et al., J. Immunol., 111(2):395-403 (1973). Monoclonal IgA antibodies have been produced and applied directly to respiratory mucosal 15 surfaces in an effort to protect against pathogen entry. Mazanec et al. J. Virol., 61:2624-2625 (1987).

Active immunization may involve challenge at the mucosal surface with intact (killed) bacteria or viruses. To avoid dangers that may be associated with this 20 approach for certain pathogens, component antigens, such as immunogenic surface components of the pathogen, are applied at a mucosal surface. In some cases, the antigens have been conjugated to larger molecules. For example, the cholera toxin B subunit has been conjugated 25 to antigens. See, Czerkinsky et al. who report oral administration of a streptococcal antigen coupled to cholera toxin B subunit in Infection and Immunity 57:1072-1077 (1989). Biodegradable microspheres have also been used as an antigen carrier. For example, 30 Eldridge et al. Curr. Top. Microbial Immunol. 146:59 et seq. (1989) report incorporation of antigen into biodegradable microspheres. The dry protein antigen is dispersed in a copolymer matrix without chemical conjugation.

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We have discovered that hydroxylated calcium phosphate (HCP) particulate is a particularly useful carrier for therapeutics to be introduced through mucosal surfaces. Conjugates of HCP and therapeutically active ingredients (e.g., biologically active substances such as antigens, or drugs) are transported across epithelium where they produce the desired therapeutic response-- e.g. they raise a poly Ig immune response.

In its most general formulation, the invention features administering an active ingredient to a mammal by applying a complex of hydroxylated calcium phosphate and the active ingredient to a mucosal surface of the mammal.

One specific embodiment of the invention generally features a method for generating antigen-sensitized Ig-A-producing lymphoblasts in a mammal. In that method, an immunogen comprising an antigen or antigen mixture in association with hydroxylated calcium phosphate (HCP) particulate is administered to a mucosal surface of the mammal. In preferred embodiments of this first aspect of the invention, the antigen-sensitized lymphoblasts are recovered and immortalized to yield an Ig-A producing hybridoma.

A second specific embodiment of the invention features a method for vaccinating a mammal (especially a human) comprising administering the above-described immunogen to a mucosal surface of the mammal.

Alternative embodiments feature delivery of therapeutics (e.g., drugs) across the mucosa.

The invention also generally features a composition of matter comprising an active ingredient in association with hydroxylated calcium phosphate particles of a size suitable for transport across epithelium.

In any of the preferred embodiments of the invention, the hydroxylated calcium phosphate is in the

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form of microparticles suitable for transport across the epithelium. Also preferably, the antigen comprises an externally available determinant of a pathogen or of spermatozoa, such as a viral coat or envelope protein, a 5 lipopolysaccharide or a cell-surface protein. One form of HCP is hydroxyapatite (HA), a commercially available crystalline hydroxylated calcium phosphate discussed below.

The preferred modes of administrations of the 10 active ingredient according to the invention are orally, vaginally, nasally, rectally, ocularly or to the middle ear. Oral administration can provide delivery to other G.I. mucosa including the intestinal mucosa.

The invention provides an efficient, polyvalent 15 complex that can adhere to the mucosa and can be transported efficiently across the epithelial barrier for its biological effect, e.g. for presentation to the mucosal immune system. Adsorption of active ingredients, particularly proteins, to HCP is relatively simple, rapid 20 and cheap, making the invention economically feasible. Moreover, HCP has a high general affinity for the antigens of interest, including proteins and other 25 antigens, making the invention broadly applicable. HCP is generally non-toxic, as evidenced by the fact that HA is an integral component of bone, and the systemic immune system routinely encounters HA during normal bone resorption, a process that occurs constantly at a microscopic level in healthy individuals. Accordingly, pure HA presumably can be safely administered without a 30 host immune response, and administration can be repeated as a vehicle for the same or different antigens, without an anti-vehicle immune response. Moreover HCP, particularly HA, is relatively inexpensive. HA can readily be reduced to a size suitable for transepithelial 35 transport by M cells; and such a reduced size is suitable

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for ingestion by macrophages and other cells of the reticuloendothelial system, so as to enhance immune response. Finally, M-cell uptake and transport of immunogens according to the invention is relatively  
5 selective.

Other features and advantages of the invention will be apparent from the following description of the preferred embodiments thereof.

Description of the Preferred Embodiments

10 Figures

Fig. 1 is a diagrammatic representation of transcytosis of antigen across M cells.

Reagents

In general, the methods and materials described  
15 below can be used as part of strategies for IgA protection which are disclosed in greater detail in a commonly owned patent application filed by Neutra, Kraehenbuhl, and Weltzin, simultaneously with this application, entitled SYNTHETIC POLY-Ig RECEPTOR,  
20 RECEPTOR-ANTIBODY COMPLEXES, PRODUCTION AND USE THEREOF, which application is hereby incorporated by reference. In particular, immunogens and methods according to the invention can be used with stabilizer protein as described in the above-referenced application to create  
25 poly Ig reagents for passive immunization.

The preferred embodiments of the invention feature hydroxyapatite-antigen conjugates and their use. Specifically, hydroxyapatite is a modified form of crystalline calcium phosphate,  $\text{Ca}_{10} (\text{PO}_4)_6 (\text{OH})_2$ . It is  
30 used as a protein fractioning reagent, due to its generalized protein-binding ability. Commercially available HA generally consists of slab-like crystals that are chemically and physically analogous to inorganic HA in normal bone tissue. As long as the starting  
35 material is free of contaminants, ingestion of HA should

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be relatively safe as evidenced by the existence of nutritional calcium/phosphorus supplements derived from ground bone, which are designed to be ingested.

Commercial high resolution HA (from Calbiochem) 5 consists of crystals varying widely in size. As provided by manufacturers, HA crystals are likely to be too large, on the whole, to efficiently cross epithelial barriers. For example, crystals over 1  $\mu\text{m}$  in length are unlikely to be taken up by M cells. Therefore, for use 10 in the invention, commercial HA crystals are broken into small, relatively uniform crystalline fragments, e.g., by sonication.

The resulting sonicated crystals vary somewhat in size but their size generally does not exceed 1  $\mu\text{m}$ . 15 Preferably a substantial portion of the HA is present as fragments of about 0.01-0.1 $\mu\text{m}$ . Fragmentation may be measured either by electron microscopy and light scattering, using standard techniques.

HA binds most proteins with high avidity and speed 20 at low or physiologically safe concentrations. Binding is thought to depend on interaction of calcium sites with acidic and phosphate groups of proteins, and interaction of phosphates with basic protein groups. Higher molecular weight proteins will tend to be more tightly 25 adsorbed to HA, but protein charge also plays a role. One gram of HA, for example, can bind about 30 mg of bovine serum albumin. Thus, HA can efficiently trap 30 protein antigen from dilute solutions without addition of crosslinking chemicals, without harsh or denaturing conditions, and without wasting valuable pure antigen.

Use

HA-adsorbed antigens can be used in active vaccination of humans or other mammals. The invention is particularly useful to protect against Pseudomonas aeruginosa, Hemophilus influenzae, Vibrio cholerae, 35

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Bordetella pertussis, Corynebacterium diphtheriae,  
Escherichia coli, Salmonella typhi and typhimurium,  
Clostridium perfringens and other enteric clostridia,  
Shigella dysenteriae, Shigella flexnerii Neisseria  
5 gonorrhoeae, Trichomonas, Entameba histolytica, Giardia  
lamblia, Streptococcus, respiratory syncytial virus,  
rotavirus, reovirus, Human Immunodeficiency Virus, Human  
T-Cell Lymphotrophic Virus, Types I and II, polio virus,  
Rhinovirus, influenza virus, herpes viruses, human  
10 papilloma virus; AIDS 2° pathogens such as Pneumocystis,  
and yeast such as monilia. The invention is also useful  
to protect against allergens that contact the respiratory  
or digestive mucosal surfaces. It is also useful to  
protect against pregnancy by cross-linking spermatozoa in  
15 the vagina, and preventing their movement through the  
cervix and uterus.

In each case, an appropriate known antigen-- e.g.  
whole pathogen or specific externally presented antigens  
such as the viral coat protein, or bacterial cell-surface  
20 proteins, pilus protein, lipopolysaccharides, viral  
capsid or envelope protein, protozoal plasma membrane  
surface component, spermatozoal surface proteins, or  
respiratory allergens--are used. Toxoids, e.g. CRM-197,  
and inactivated diphtheria toxin reported by Uchida et  
25 al. J. Biol. Chem. 248:3838-3844 (1973) may be used. The  
antigen is used according to the above procedure to  
generate hybridomas secreting the desired protective  
antibodies. As just a few examples, WO88/08437 (hereby  
incorporated by reference) discloses a tcPA pilus protein  
30 suitable for forming anti-V. cholerae monoclonal poly-Ig  
antibodies. U.S. Pat. 4,725,669 discloses the HIV  
(HTLV-III) envelope glycoprotein suitable for forming  
anti-HIV poly-Ig monoclonal antibodies. The following  
patents and patent applications disclose preparation of  
35 antigens for protection against Streptococcal infections,

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particularly infection by Group B Streptococcus: U.S. Pat. 4,367,221; 4,367,222; 4,367,223; 4,356,263; 4,207,414 (RE 31,672); and WO 87/06267.

Other suitable antigens are disclosed in Bacterial Vaccines and Local Immunity Proceedings of the Sclavo International Conf., Siena, Italy 17-19 November 1986. Specific reagents suitable for HIV antigens are disclosed in AIDS Research and Reference Reagent Program, National Institute of Health, June 1989. For example, gp 120 is sold by MicroGeneSys, Inc. Spermatozoa cell-surface antigens such as LDH-C4 are also known. See, e.g. Shaha et al. and Talwar et al. Vaccine 7:97-100 (1989); and Shaha et al. Int. J. Androl. 11:479 (1988).

Of particular significance in the selection of antigens for practice of the invention is that mucosal protection involves cross-linking to prevent entry into the body, and this mechanism does not require that the polymeric antibody kill or "neutralize" the pathogen. In contrast, systemic (IgG) protection involves binding which, to be effective, generally must neutralize the pathogen. Thus, not every IgG antibody which binds to the pathogen is protective, as illustrated by the existence of monoclonal antibodies that specifically bind Vibrio cholerae, but do not neutralize it in the sense of preventing it from colonizing, growing and manifesting clinical symptoms in its host. The universe of antigens and determinants available to raise protective IgA antibodies is thus significantly increased.

Specifically, the HA-adsorbed antigen is prepared according to the method outlined above with appropriate modifications for production in bulk. The HA-adsorbed antigen or antigen mixture is compounded in a physiologically acceptable vehicle, and applied directly to or delivered to the mucosal surface tissue, e.g., to oral, nasal, rectal and/or vaginal surfaces. The

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preparation is administered in an aerosol, a suspension, a capsule and/or a suppository. Those skilled in the art will understand how to formulate such vehicles by known techniques.

5 Use

The invention is also particularly useful for manufacture of IgA-producing hybridomas. Such hybridomas are readily produced by challenging a mammal, e.g., by applying the above described composition to a mucosal 10 surface, and then recovering a lymphoid cell from Peyer's patch mucosa or other mucosa which are rich in lymphoid tissue and then fusing the lymphoid cell to a myeloma cell by known techniques. See, e.g., the above-referenced commonly owned, simultaneously filed, 15 U.S. patent application USSN 07/510,161.

The following examples are provided to illustrate, but not to limit the invention.

Example 1

2 gm HA (Calbiochem) suspended in 20 ml PBS is 20 sonicated with a probe sonicator for 30 minutes at room temperate, using the high setting (140, 80% duty cycle, of a Microson cell disrupter (Heat Systems Ultrasonics, Inc., Farmingdale, NY). Average crystal size is approximately  $0.01 \times 0.1 \mu\text{m}$  after sonication, as measured 25 by electron microscopy. Adequate sonication can be monitored by spectrophotometric absorbance. For example, absorbance (read at 650 nm) of a 2 mg/ml suspension before sonication was 0.108 O.D. units and after reduction of crystal size (as above), O.D. increased to 30 2.060.

Example 2

HA crystals of this size, coated to saturation with proteins of various sizes ranging from 65 kD (albumin) to 450 kD (ferritin), when introduced into the 35 intestinal lumen of mice and rabbits, were selectively

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bound and transported by M cells of Peyer's patch. This method effectively delivers proteins to cells of the mucosal immune system, in a particulate immunogenic form.

For oral immunization of a 25-gram mouse: a small  
5 amount (e.g., 30 µg) of protein in 200 µl PBS is mixed  
with an appropriate amount (e.g., 1 mg of HA for 30 µg  
protein) of presonicated and washed HA. The mixture is  
agititated for 1 hr. at 4°C, and HA is then pelleted by  
spinning for 2 min. at 10,000 rpm in a microfuge. The  
10 pellet is resuspended by vortexing or sonicating in 200  
µl of water which may contain a buffer to neutralize  
gastric acid.

Other embodiments are within the following claims.

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Claims

1        1. A method for administering an active  
2 ingredient to a mammal comprising administering a complex  
3 to a mucosal surface of the mammal, said complex  
4 comprising said active ingredient in association with  
5 hydroxylated calcium phosphate.

1        2. The method of claim 1 in which the  
2 hydroxylated calcium phosphate is present in the form of  
3 particles suitable for transport across epithelium.

1        3. The method of claim 1 in which the complex is  
2 an immunogen comprising an antigen or antigen mixture in  
3 association with hydroxylated calcium phosphate.

1        4. The method of claim 3 in which the  
2 administration of the complex generates IgA-producing  
3 lymphoblasts in the mammal which are sensitized to the  
4 antigen.

1        5. The method of claim 3 in which the  
2 administration of the immunogen vaccinates the mammal,  
3 protecting it against mucosally introduced agents that  
4 present the antigen.

1        6. The method of claims 3, 4 or 5 in which said  
2 antigen comprises an externally available determinant of  
3 a pathogen or of spermatozoa.

1        7. The method of claims 3, 4 or 5 in which said  
2 antigen is a viral coat protein, a viral envelope  
3 protein, a cell-surface lipopolysaccharide, or a  
4 cell-surface protein.

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1        8. The method of claims 3, 4 or 5 in which said  
2 immunogen is administered orally, vaginally, nasally,  
3 rectally, ocularly, or to the middle ear.

1        9. The method of claim 4 further comprising  
2 isolating and immortalizing a plurality of lymphoblasts  
3 from said mammal and identifying among said immortalized  
4 lymphoblasts at least one immortalized lymphoblast that  
5 secretes IgA that is capable of specifically binding to  
6 said antigen.

1        10. A composition of matter comprising an active  
2 ingredient in association with hydroxylated calcium  
3 phosphate microparticles of a size suitable for transport  
4 across epithelium in association with the active  
5 ingredient.

1        11. The composition of claim 10 wherein said  
2 complex is an immunogen for raising a mucosal immune  
3 response in a mammal, and said active ingredient  
4 comprises an antigenic determinant to which an immune  
5 response is desired.

1        12. The immunogen of claim 11 wherein said  
2 antigenic determinant is an externally available  
3 determinant of a pathogen or of spermatozoa.

1        13. The immunogen of claim 12 wherein said  
2 antigenic determinant is a determinant of viral coat  
3 protein, a viral envelope protein, a cell-surface  
4 lipopolysaccharide, or a cell-surface protein.

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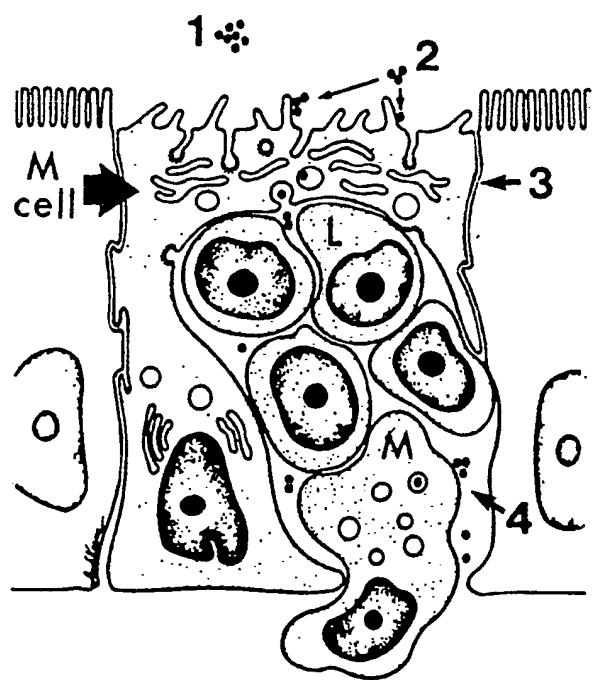


FIG. I

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# INTERNATIONAL SEARCH REPORT

International Application No PCT/US91/02599

<b>I. CLASSIFICATION OF SUBJECT MATTER</b> (if several classification symbols apply, indicate all)		
According to International Patent Classification (IPC) or to both National Classification and IPC IPC (5): A61K 39/12, 31/66, 39/385, 37/16; A01N 37/18 US.C1: 424/603, 88, 89; 514/7, 2		
<b>II. FIELDS SEARCHED</b>		
Minimum Documentation Searched *		
Classification System	Classification Symbols	
U.S. Cl: 424/603, 88, 89; 514/7, 2.		
Documentation Searched other than Minimum Documentation to the Extent that such Documents are Included in the Fields Searched *		
<b>III. DOCUMENTS CONSIDERED TO BE RELEVANT</b> †		
Category *	Citation of Document, †† with indication, where appropriate, of the relevant passages ‡‡	† Relevant to Claim No. ††
A	F.P. A, 0,175,286 (MCW-RES. found) 26 March 1986, see abstract.	1, 3, 4
A	JP. A. 63.196.281 (Sumitomo Elec. 15 August 1988, see abstract.	10-13
A	JP. A, 59.101.145 (Showa) 11 June 1984, see abstract.	1-13
Y	US. A, 4,722,840 (Valenzuela et al) 02 February 1988, see entire document.	1-13

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## IV. CERTIFICATION

Date of the Actual Completion of the International Search

13 June 1991

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Date of Issue of the International Search Report

17 JUL 1991



Lila Feisee

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